



SwisensPoleno: Real-time assessment of *P. infestans* sporangia in the air

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Content

- The measuring system SwisensPoleno and goal of the project
- Development of the *P. infestans* specific classifier
- Field trial setup and Lab trials
- Results
- Conclusions, Challenges and Outlook

Automatic Real-time Monitoring of Bioaerosol

Current method for identifying pollen and spores



Sampling



Visualization

- **Manual identification**
- **Several days delay**

New method:



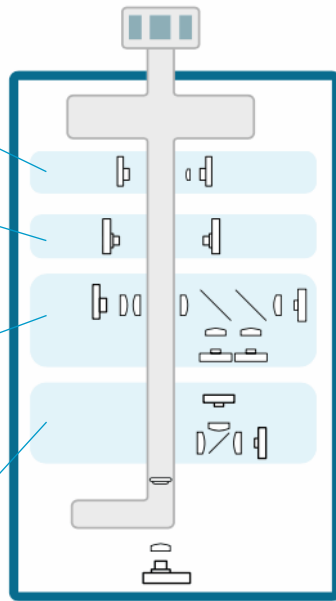
- **Fully automatic**
- **Continuous monitoring**
- **Real time identification**



SwisensPoleno

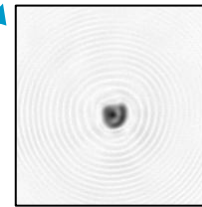
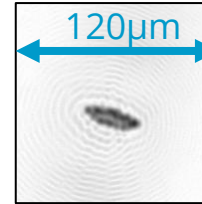
SwisensPoleno Jupiter

1. Trigger, scattering
2. Holographic imaging
3. Fluorescence spectra and lifetimes
4. Polarized time-resolved scattered light

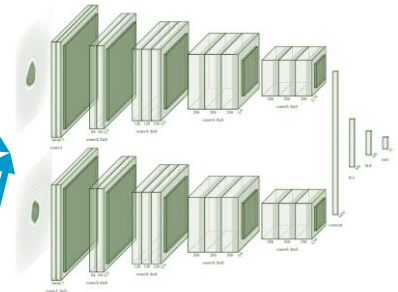


Holographic images provide rich **morphological** data

Classification in real-time classification using Machine Learning



Relative fluorescence spectra provide **fingerprints** on the **chemical composition**



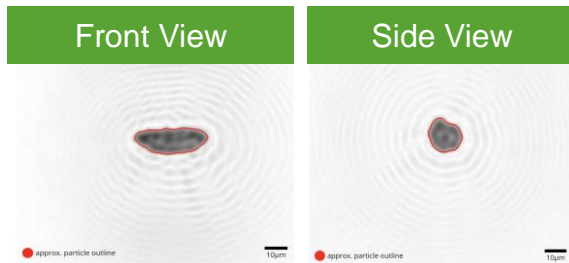
Prediction result

0.0	Water droplets
0.0	<i>Alternaria alternata</i>
0.0	<i>Alternaria solani</i>
0.0	<i>Erysipe necator</i>
0.9	<i>Phytophthora infestans</i>
0.1	<i>Plasmopara viticola</i>
0.0	<i>Alnus Glutinosa</i>
0.0	<i>Betula Pendula</i>
0.0	<i>Carpineus sp.</i>
0.0	<i>Fraxineus excelsior</i>

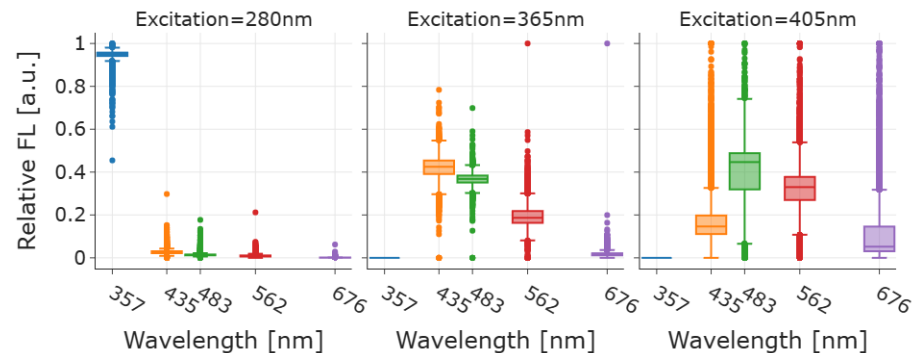


Development of classifier

- The classifier (Holo-FL 2024) was trained on 443'874 measurement events to distinguish between 58 different classes (21 spore taxa, 35 pollen taxa, 2 others)
- Fluorescence spectra data was included to improve classification of morphologically similar particles
- Empirical thresholds based on collected datasets are applied after classification



Relative Fluorescence Spectra of 11'146 *P. infestans* spores

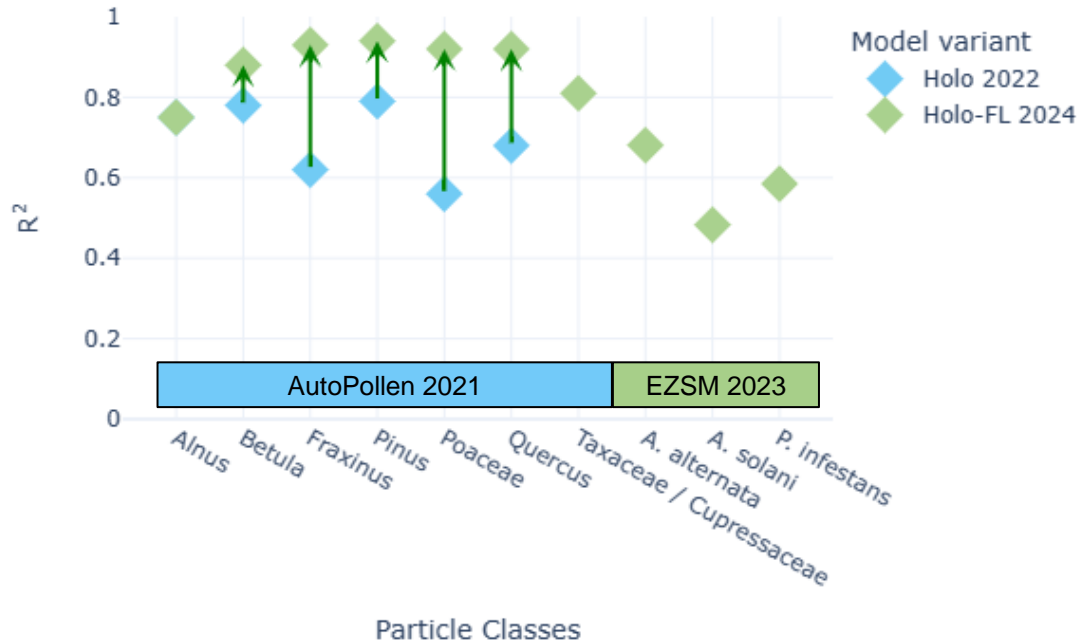




Classifier improvements

Holo 2022 <> Holo-FL 2024

- The new classifier using fluorescence data significantly improves the performance of particle classification for pollen (AutoPollen 2021)
- Spores and sporangia relevant for potato farming correlate well with reference Hirst data.
 - *P. infestans* R^2 : 0.59
 - *A. alternata* R^2 : 0.68
 - *A. solani* R^2 : 0.483



Field and Lab trails

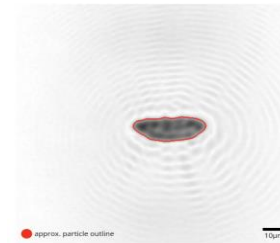
Field:

- untreated potato plots in Zurich and Zollikofen (BE)
- SwisensPoleno Jupiter (automatic) and traditional Hirst spore trap (manually)
- Late blight assessment (occurrence, spread) as well as registration of **main infection** and **sporulation periods** (MISP PhytoPRE)



Lab:

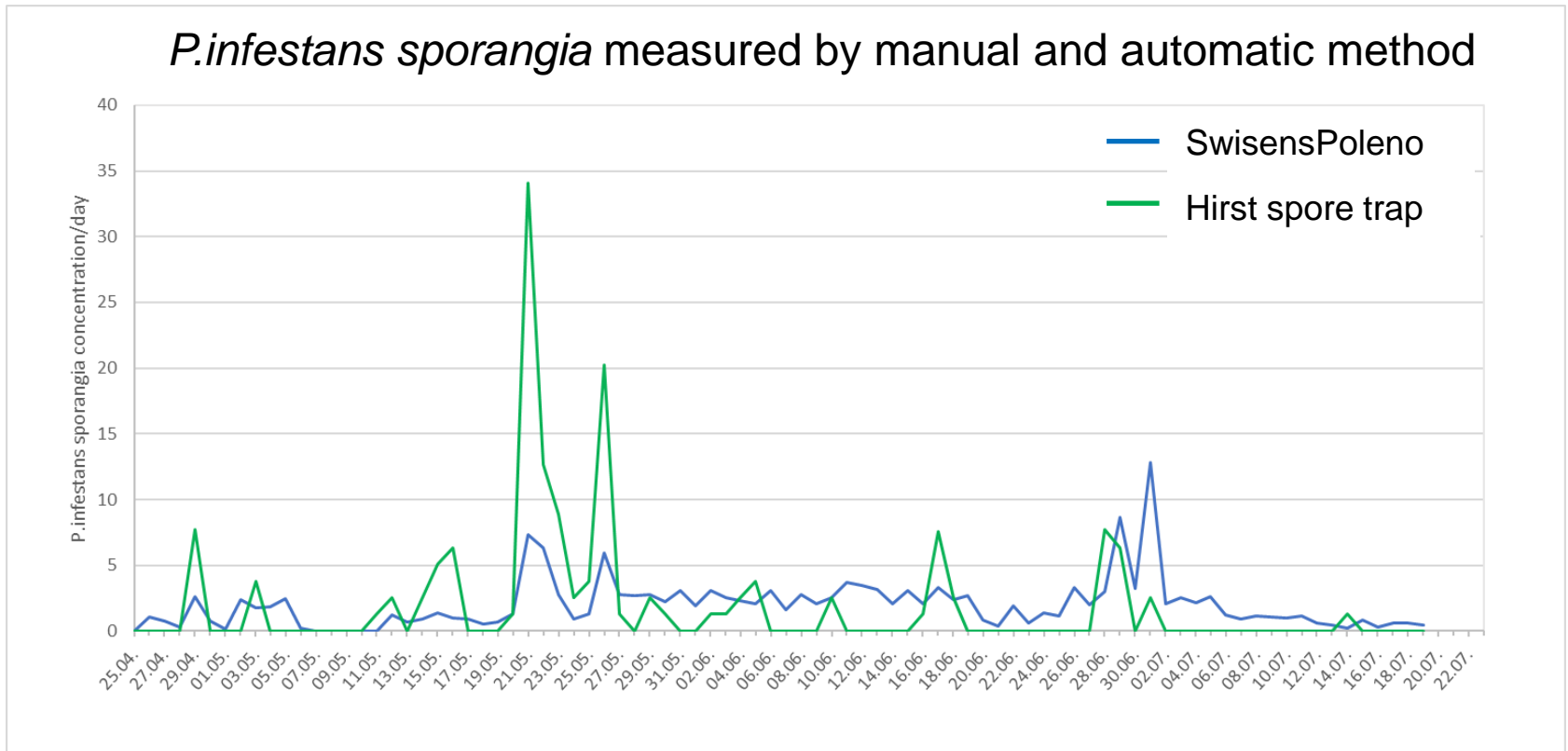
- clean datasets for classifier development
- SwisensPoleno system in the lab
- Measurements with artificially and naturally infected leaves/tubers, different Pi isolates





Relation between measured sporangia concentration by different spore traps

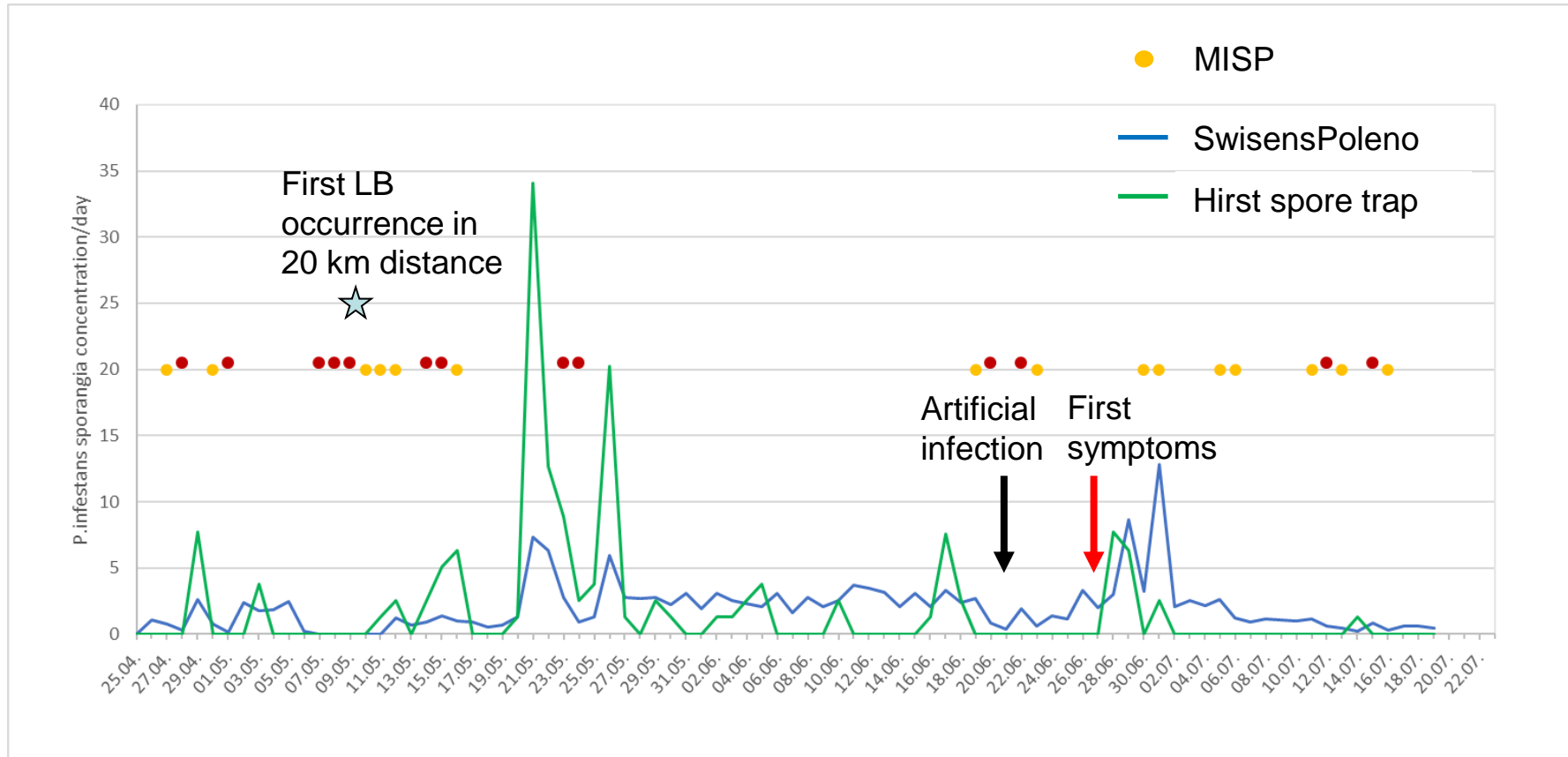
Location: Zurich Reckenholz, 2023



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Relation between assessed DSS-infection risk, LB occurrence and sporangia concentration

Location: Zurich Reckenholz, 2023

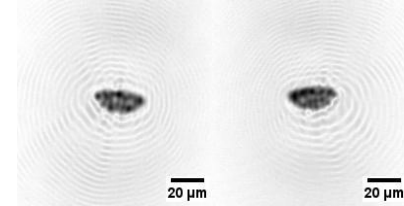


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Conclusions

- ✓ Detection and identification of *P. infestans* sporangia successful with SwisensPoleno Jupiter system

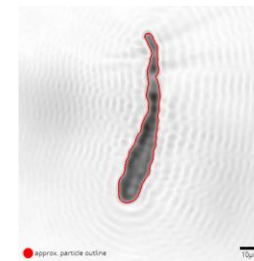


- ✓ Correlation between SwisensPoleno Jupiter, Hirst spore trap and occurrence of LB in the field

Reliable real-time measurement with SwisensPoleno Jupiter based on the specific developed classifier for *P. infestans* has to be validated with more sites and seasons

🇨🇭 Challenges and Outlook

- Identification of the minimal spores concentration in the air which leads to a successful *P. infestans* infection in the field
- Analysis of weather data
- Do we find *P. infestans* sporangia also at higher elevations (rooftop) and can these measurements be used for reliable assessments?
- How to integrate into DSS (PhytoPRE) ?
- Further field trials without artificial infection to validate the classifier at several locations
- Expansion of identifiable spores, especially *Alternaria solani* and *A. alternata*



A. solani



A. alternata



Thank you for your attention



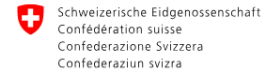
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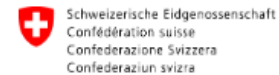


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Question we would like to discuss with you

- Where do you see value of automatic spore monitoring?
- How would you quantify the value of spore detection?
- How dense should the measurement network be?